

Population Screening Programme of Thalassaemic Patients in Barpeta Region of Assam

Anirban Roy Chowdhury¹, Sudipa Chakravarty², Shakuntala Ghose³, Amit Chakravarty⁴

¹*Department of Medical Lab Technology, Institute of Genetic Engineering. 30 Thakurhat Road. Badu. Kolkata- 700128, West Bengal, India*

^{2,4}*Director, Inheritance Healthcare, A Unit of SAG Foundation, Agate Akash Appt, Flat No. 1D, Block A, 1st Floor, Thakdari Panchayat Road, Action Area I, Newtown, West Bengal 700156, India*

³*Mg. Director, Dept of Haematology, Repose Clinic & Research Centre Pvt.Ltd. (Empanelled under QUALITY COUNCIL OF INDIA). 20C, Ustad Bade Golam Ali Khan Sarani (Broad Street). Ward No.65.PO-Ballygunge.PS-Karaya.Kolkata-700019. (West Bengal). India*

Abstract—The β -thalassemias are characterized by a very heterogeneous group of inherited mutations causing abnormal expression of globin genes, leading to total absence or quantitative reduction of synthesis of β -globin chains (1–3). This disease is frequent in the Mediterranean area, Middle East, Africa and Asia. More than 200 different mutations have been identified in β -thalassaemia patients, including deletions of the β -gene region, stop codons leading to premature termination of a non-functional β -globin chain, mutations suppressing correct maturation of the β -globin RNA precursor, most of all need regular blood transfusions.

In this present study we performed a population screening programme of thalassaemic patients in Barpeta region of Assam. In this programme we found 82 E-Bata, 18 Beta among 100 patients.

Index Terms— β -Thalassaemia, β -globin chain, Blood transfusion.

I. INTRODUCTION

The β -thalassemias are characterized by a very heterogeneous group of inherited mutations causing abnormal expression of globin genes, leading to total absence or quantitative reduction of synthesis of β -globin chains (1–3). This disease is frequent in the Mediterranean area, Middle East, Africa and Asia. More than 200 different mutations have been identified in β -thalassaemia patients, including deletions of the β -gene region, stop codons leading to premature termination of a non-functional β -globin chain, mutations suppressing correct maturation of the β -globin RNA precursor, most of all need regular blood transfusions. (1–3,4,5).

It is firmly established that increasing the production of γ -globin leads to a decrease in the imbalance between β and non- β -chains and the consequent reduction of haemolysis. (6–9), including the very interesting finding that a group of genetic mutations, known as hereditary persistence of fetal hemoglobin (HPFH), are associated with high levels of HbF in adults (10–13) resulting in a mild phenotype. Most of the HPFH patients homozygous for β -thalassaemia do not need blood transfusion. Therefore, there has been considerable interest in recent years to solve this problem in finding ways of increasing production of HbF and reactivation of the γ -globin genes (14–23).

II. MATERIALS AND METHODS

A. Study groups:

Patients with HPLC-screened documented Beta thalassaemia, HbE-beta thalassaemia genotypes have been considered in this screening programme. Total 100 patients were evaluated in this screening programme.

III. RESULT

In our population screening study, we evaluated total 100 patients. Among which 82 patients with Hb-E-beta and 18 patients with Beta thalassaemia were observed. This was clearly depicted in Chart 1. We also evaluated the male and female percentage among Beta and Hb-E-beta Thalassaemic patients. Which were clearly depicted in Chart 2 and Chart 3

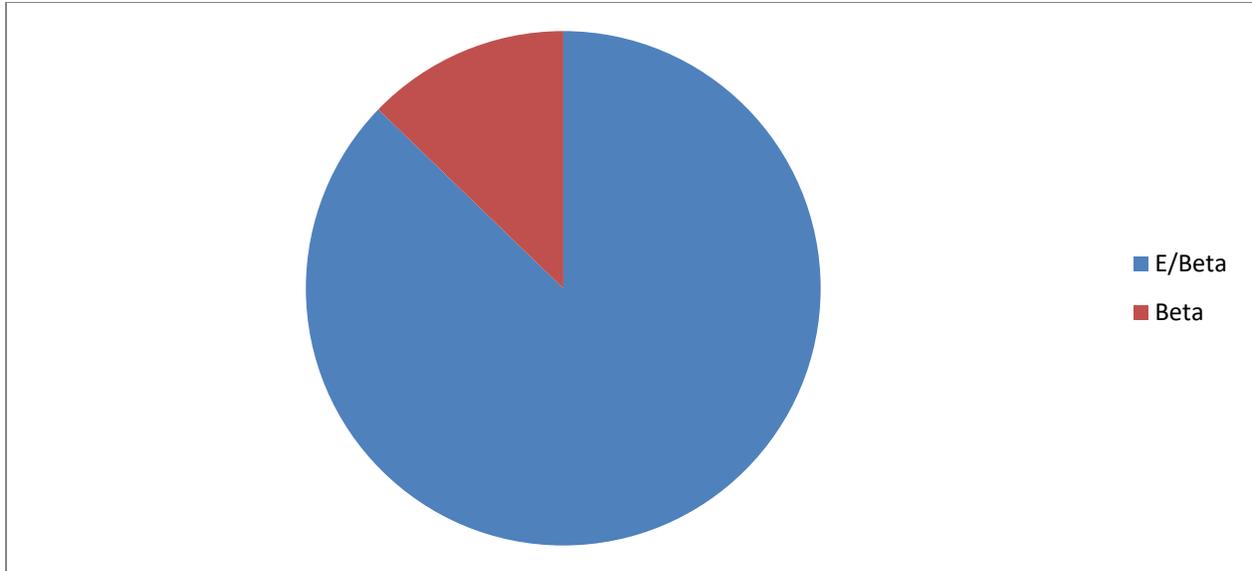


Chart 1: Population Screening of Beta, E-Beta, among total 100 thalassaemic patients in Barpeta region of Assam.

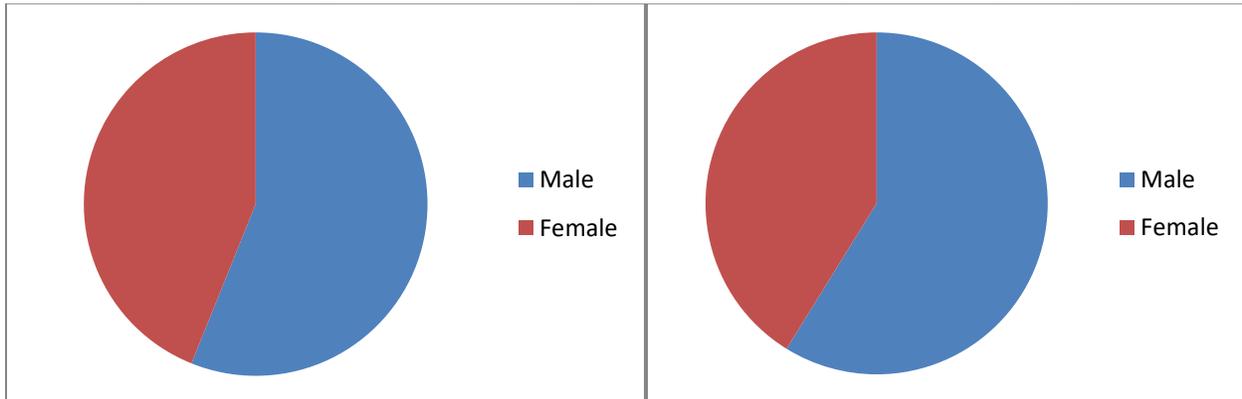


Chart 2 Male and female percentage among 82 E-Beta Thalassaemic Patients.

Chart 3 Male and female percentage among 18 Beta Thalassaemic Patients.

IV. DISCUSSION

β -thalassemias (β -thal) are common inherited red cell disorders characterized by absent or reduced synthesis of β -globin chains. Despite extensive knowledge of the molecular defects causing β -thalassaemia, less is known about the mechanisms responsible for the associated ineffective erythropoiesis and reduced red cell survival (24-31). Increased levels of reactive oxygen species (ROS) have been reported to contribute to the anemia of β -thalassaemia, although the effects of ROS have not been fully defined (24, 26-31). Exogenous anti-oxidant molecules might represent complementary therapeutic strategies to

counteract the toxic effects of ROS in β -thalassaemia. However, few of them have been shown to beneficially affect *in vivo* β -thalassaemic red cell features and/or thalassaemic ineffective erythropoiesis *in vivo*. (27-31).

In our present population screening study, we observed that the occurrence of E-beta thalassaemia was very higher than Beta thalassaemia in Barpeta region of Assam which was clearly mentioned in chart 1 and other two charts, Chart 2 and 3 we observed that both E-Beta and Beta cases the effected males were always higher then effected females.

V. CONCLUSION

Beta Thalassemia is an inherited disorder in which either very few or no red blood cells are produced by the bone marrow after infancy. The treatment is monthly whole blood transfusions and the use of a drug which is extremely toxic and cannot be used with children. The disease dramatically impacts the sufferers' quality of life and often results in death around the age of puberty. Because it is more common in less developed countries where it is virtually impossible for anyone other than the very wealthy to obtain regular supplies of clean whole blood for the required transfusions, the fatality rate is high. Even if the patient is able to obtain monthly transfusions and is able to afford the drugs to treat the disease, he or she is constantly anemic and lacking of energy. In our present study we conclude that in Assam BARPETA region the occurrence of E-Beta thalassaemia was higher than Beta Thalassaemia and the percentage of affected male patients were higher than the affected female patients.

A. Funding statement:

This research received no specific grant from any funding agency in the public, commercial or not-for-profit sectors. All the research work done by the affiliated institution funding.

B. Competing Interests Statement:

The authors declare that they have no competing interests.

C. Data Sharing Statement:

We cannot share any unpublished data with other laboratory or person.

D. Patients Consent Statement:

The signed consent from all the patients were taken before test was performed and kept them as official documents. In case of any unusual condition it will be presented in front of the concerned person.

VI. ACKNOWLEDGEMENT

Author acknowledges Institute of Genetic Engineering for funding and affiliation. We are also thankful to Repose Clinic & Research Centre Pvt.Ltd. and other associated persons of IGE for their enthusiastic participation.

REFERENCES

- [1] Steinberg MH, Forget BG, Higgs DR, Nagel RL. Disorders of Hemoglobin: Genetics, Pathophysiology and Clinical Management. Cambridge, UK: Cambridge University Press, 2001.
- [2] Thein SL. Genetic insights into the clinical diversity of beta thalassaemia. *Br J Haematol* 2004; 124:264–74.
- [3] Old JM. Screening and genetic diagnosis of haemoglobin disorders. *Blood Rev* 2003; 17:43–53.
- [4] Wenzel E, Somoza V. Metabolism and bioavailability of trans-resveratrol. *Mol Nutr Food Res.* 2005;49(5):472-481. (PubMed)
- [5] Goldberg DM, Yan J, Soleas GJ. Absorption of three wine-related polyphenols in three different matrices by healthy subjects. *Clin Biochem.* 2003;36(1):79-87. (PubMed)
- [6] Osti F, Corradini FG, Hanau S, Matteuzzi M, Gambari R. Human leukemia K562 cells: induction to erythroid differentiation by guanine, guanosine and guanine nucleotides. *Haematologica* 1997; 82:395–401.
- [7] Lampronti I, Bianchi N, Zuccato C, Medici A, Bergamini P, Gambari R. Effects on erythroid differentiation of platinum (II) complexes of synthetic bile acid derivatives. *Bioorg Med Chem* 2006; 14:5204–10.
- [8] Olivieri NF. Reactivation of fetal hemoglobin in patients with betathalassaemia. *Semin Hematol* 1996; 33:24–42.
- [9] Blau CA, Stamatoyannopoulos G. Hemoglobin switching and its clinical implications. *Curr Opin Hematol* 1994; 1:136–42.
- [10] Forget BG. Molecular basis of hereditary persistence of fetal hemoglobin. *Ann N Y Acad Sci* 1998; 850:38–44.
- [11] Bhardwaj U, McCabe ER. Multiplex-PCR assay for the deletions causing hereditary persistence of fetal hemoglobin. *Mol Diagn* 2005; 9:151–6.
- [12] Liu LR, Du ZW, Zhao HL, Liu XL, Huang XD, Shen J, et al. T to C substitution at -175 or -173 of the gamma-globin promoter affects GATA-1 and Oct-1 binding in vitro differently but can independently reproduce the hereditary persistence of fetal hemoglobin phenotype in transgenic mice. *J Biol Chem* 2005; 280:7452–9.
- [13] Garner C, Dew TK, Sherwood R, Rees D, Thein SL. Heterocellular hereditary persistence of fetal

- haemoglobin affects the haematological parameters of beta-thalassaemia trait. *Br J Haematol* 2003; 123:353–8.
- [14] Lal A, Vichinsky E. The role of fetal hemoglobin-enhancing agents in thalassemia. *Semin Hematol* 2004; 41:17–22.
- [15] Rodgers GP, Dover GJ, Uyesaka N, Noguchi CT, Schechter AN, Nienhuis AW. Augmentation by erythropoietin of the fetal hemoglobin response to hydroxyurea in sickle cell disease. *N Engl J Med* 1993; 328:73–80.
- [16] Rodgers GP, Rachmilewitz EA. Novel treatment options in the severe beta-globin disorders. *Br J Haematol* 1995; 91:263–8.
- [17] Steinberg MH, Lu LZ, Barton FB, Terrin ML, Charache S, Dover GJ. Fetal hemoglobin in sickle cell anemia: determinants of response to hydroxyurea. Multicenter Study of Hydroxyurea. *Blood* 1997; 89:1078–88.
- [18] Olivieri NF, Rees DC, Ginder GD, Thein SL, Wayne JS, Chang L, et al. Elimination of transfusions through induction of fetal hemoglobin synthesis in Cooley’s anemia. *Ann N Y Acad Sci* 1998; 850:100–9.
- [19] Swank RA, Stamatoyannopoulos G. Fetal gene reactivation. *Curr Opin Genet Dev* 1998; 8:366–70.
- [20] Cao H. Pharmacological induction of fetal hemoglobin synthesis using histone deacetylase inhibitors. *Hematology* 2004; 9:223–33.
- [21] Lo L, Singer ST. Thalassemia: current approach to an old disease. *Pediatr Clin North Am* 2002; 49:1165–91.
- [22] Atweh GF, Loukopoulos D. Pharmacological induction of fetal hemoglobin in sickle cell disease and beta-thalassemia. *Semin Hematol* 2001; 38:367–73.
- [23] Olivieri NF, Weatherall DJ. The therapeutic reactivation of fetal haemoglobin. *Hum Mol Genet* 1998; 7:1655–8.
- [24] De Franceschi L, Bertoldi M, De Falco L, Santos Franco S, Ronzoni L, Turrini F, et al. Oxidative stress modulates heme synthesis and induces peroxiredoxin-2 as a novel cytoprotective response in beta-thalassemic erythropoiesis. *Haematologica*. 2011;96(11): 1595–604 [PMC free article][PubMed]
- [25] Rund D, Rachmilewitz E. Beta-thalassemia. *N Engl J Med*. 2005;353(11):1135–46 [PubMed]
- [26] de Franceschi L, Turrini F, Honczarenko M, Ayi K, Rivera A, Fleming MD, et al. In vivo reduction of erythrocyte oxidant stress in a murine model of beta-thalassemia. *Haematologica*. 2004;89(11):1287–98 [PubMed]
- [27] De Franceschi L, Ronzoni L, Cappellini MD, Cimmino F, Siciliano A, Alper SL, et al. K-CL co-transport plays an important role in normal and beta thalassemic erythropoiesis. *Haematologica*. 2007;92(10):1319–26 [PubMed]
- [28] Olivieri O, De Franceschi L, Capellini MD, Girelli D, Corrocher R, Brugnara C. Oxidative damage and erythrocyte membrane transport abnormalities in thalassemias. *Blood*. 1994; 84(1):315–20 [PubMed]
- [29] Ginzburg Y, Rivella S. β -thalassemia: a model for elucidating the dynamic regulation of ineffective erythropoiesis and iron metabolism. *Blood*. 2011;118(16):4321–30 [PMC free article][PubMed]
- [30] Liu J, Zhang J, Ginzburg Y, Li H, Xue F, De Franceschi L, et al. Quantitative analysis of murine terminal erythroid differentiation in vivo: novel method to study normal and disordered erythropoiesis. *Blood*. 2013;121(8): e43–9 [PMC free article][PubMed]
- [31] Fibach E, Rachmilewitz E. The role of oxidative stress in hemolytic anemia. *Curr Mol Med*. 2008;8(7):609–19 [PubMed]