In Vitro Anticataract Activity of Tamarindus Indica Linn, Againest Glucose-Induced Cataractogeisins

Thaware Sachin Bhaskarrao¹, Tarkase A S², Salgar P A ³

1,2,3</sup> Aditay Pharmacy College, Beed

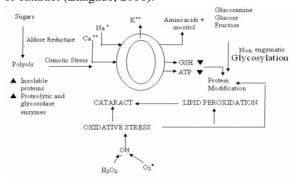
Abstract— These instructions give you guidelines for preparing papers for the International Journal of Innovative Research in Technology (IJIRT). Use this document as a template if you are using Microsoft Word 6.0 or later. Otherwise, use this document as an instruction set. The electronic file of your paper will be formatted further at International Journal of Innovative Research in Technology. Define all symbols used in the abstract. Do not cite references in the abstract. Do not delete the blank line immediately above the abstract.

Index Terms— About four(minimum) key words or phrases in alphabetical order, separated by commas.

I. INTRODUCTIO)

The crystalline lens is a biconvex, vascular, transparent structure enclosed by a capsule, a basement membrane secreted by the lens epithelium. The capsule, responsible for moulding the lens substance during accommodation, is thickest in the equatorial zone and thinnest at the posterior pole of the lens. The normal lens is transparent and any congenital or acquired opacity in the lens capsule or substance, irrespective of the effect on vision, is a cataract (Kanski, 2003).

Cataract is the opacification of lens often associated with old age and is a major complication of diabetes mellitus because higher glycosylated hemoglobin levels are significantly associated with increased risk of cataract (Langade, 2006).



Mechanisms associated with cataract

There are mainly two mechanisms involved in the development of cataract. Trypotophan which is an amino acid is present in eye lens in higher concentration. When it absorbs UV radiation, it forms N-Formyl Kynurenine. It may combine with the 3-OH Kynurenine and riboflavin which are lens photosensitizers, they absorb photons form the light and emits electrons. These electrons react with the molecular O₂ to form super oxide anion radical which reacts with Na⁺ K⁺ ATPase pump in eye and may cause the swelling of the eye and cause lens opacification.

II. PROCEDURE

Preparation of extraction

The fresh leaves of *Tamarindus indica* Linn are collected, dried in shade under room temperature, powdered mechanically and sieved through No. 20 mesh sieve. The finely powdered leaves were kept in an airtight container until the time of use. The extraction was carried out by continuous hot percolation method using Soxhlet apparatus. The solvent used was a mixture of methanol: water in the ratio of 7:3. About 100 g of powder was extracted with 600 ml of solvent. The extract was concentrated to dryness under controlled temperature between 40-50 °C.

Preparation of lens homogenate

After 72 h of incubation, homogenate of lenses (10% w/v) was prepared in Tris buffer (0.23 mM, pH 7.8) containing 0.25x10⁻³ M EDTA. The homogenate was centrifuged at 10,000 g for 1 h and the supernatant was used for estimation of total protein (TP), determination of the end products of lipid peroxidation namely malondialdehyde (MDA) and lipid hydroperoxides (LH), enzymatic antioxidants like superoxide dismutase (SOD), catalase (CAT), glutathione

reductase (GSSH), peroxidase (Px), and glutathione peroxidase (GPx), and the nonenzymatic antioxidant reduced glutathione (GSH).

Determination of enzymatic antioxidants Estimation of superoxide dismutase (SOD)

SOD activity was determined by the inhibition of auto catalyzed adrenochrome formation in the presence of the homogenate at 480 nm. The reaction mixture contained 150 µl of lens homogenate, 1.8 ml of carbonate buffer (30 mM, pH 10.2), and 0.7 ml of distilled water and 400 µl of epinephrine (45 mM). Auto oxidation of epinephrine to adrenochrome was performed in a control tube without the homogenate (Kakkar *et al.*, 1984).

IV. UNITS

Estimation of glutathione reductase (GSSH)

The activity enzyme was determined spectrophotometrically by the decrease in absorbance of NADPH at 340 nm. The reaction mixture contained 2.1 ml of 0.25 mM, potassium phosphate buffer pH 7.6, 0.1 ml of 0.001 M NADPH, 0.2 ml of 0.0165 M oxidized glutathione and 0.1 ml (10 mg/ml) of bovine serum albumin (BSA). The reaction was started by the addition of 0.02 ml of tissue homogenate with mixing and the decrease in the absorbance at 340 nm was measured for 3 min against a blank. Glutathione reductase activity was expressed as nmoles NADPHoxidized /min/mg lens protein at 30°C (Racker, 1955).

Estimation of peroxidase (Px)

Peroxidase activity was measured spectrophotometrically by following the change in absorbance at 460 nm due to O-dianisidine oxidation in the presence of H₂O₂ and enzyme. Reaction mixture contained 0.2ml of 15 mM O-dianisidine, 0.1ml of tissue homogenate and 2.5 ml of 0.1M potassium phosphate buffer pH 5.0 and were incubated at 37°C for 15 min and the reaction was started with the addition of 0.2 ml of hydrogen peroxide and the absorbance at 460 nm was followed against a blank, spectrophotometrically for about 3-5 min at 37°C. Unit of enzyme activity defined as umoles of Odianisidine/min at 37°C (Lobarzewski and Ginalska, 1995).

V. HELPFUL HINTS

Glutathione peroxidase activity was measured by the procedure of Paglia and Valentine, 1967. The reaction mixture consists of 0.2 ml of 0.4 M Tris buffer, 0.1 ml of sodium azide, 0.1 ml of hydrogen peroxide, 0.2 ml of glutathione and 0.2 ml of supernatant incubated at 37°C for 10 min. The reaction was arrested by the addition of 10 % TCA and the absorbance was measured at 340 nm. Activity was expressed as nmoles/min/mg lens protein.

The method was based on the reaction of reduced glutathione with dithiobisnitrobenzoic acid (DTNB) to give a compound that absorbs at 412 nm. To the homogenate, 0.1 ml of 10% TCA was added and centrifuged. About 0.1 ml of supernatant was treated with 0.5 ml of Ellmans reagent (19.8 mg of 5, 5'-dithiobisnitro benzoic acid DTNB in 100 ml of 0.1% sodium nitrate) and 3.0 ml of phosphate buffer (0.2 M, pH 8.0) and the absorbance was read at 412nm. Activity was expressed as nmoles/min/mg lens protein (Ellman, 1959).

VI. PUBLICATIONPRINCIPLES

The contents of the journal are peer-reviewed and archival. International Journal of Innovative Research in Technology publishes scholarly articles of archival value as well as tutorial expositions and critical reviews of classical subjects and topics of current interest.

Authors should consider the following points:

- Technical papers submitted for publication must advance the state of knowledge and must cite relevant prior work.
- 2) The length of a submitted paper should be commensurate with the importance, or appropriate to the complexity, of the work. For example, an obvious extension of previously published work might not be appropriate for publication or might be adequately treated in just a few pages.
- Authors must convince both peer reviewers and the editors of the scientific and technical merit of a paper; the standards of proof are higher when extraordinary or unexpected results are reported.
- 4) Because replication is required for scientific progress,

Papers submitted for publication must provide sufficient information to allow reader stoper form similar experiments or calculations and use the reported results. Although not everything need be disclosed, a paper must contain new, useable, and fully described information. For example, as pecimen's chemical composition need not be reported if the main purpose of a paper is to introduce a new measurement technique. Authors should expect to be challenged by reviewers if the results are not supported by adequated at a and critical details.

VII. CONCLUSION

The present thesis entitled "In vitro anticataract activity of Tamarindus indica L. against glucose-induced cataractogenesis" deals with the exploration of pharmacological and phytochemical screening of the selected Indian medicinal plant Tamarindus indica L. belonging to the family Fabaceae, which is traditionally used by the local people and tribals in India for the treatment of inflammatory swelling, tumors, ringworm; useful in disease of blood, small pox, ophthalmia and other eye diseases, earache, snake-bike (Krithikar and Basu, (1981).

Cataract is one of the leading causes of visual disability often leading to blindness. It is an agerelated phenomenon over and above, oxidative stress also plays an important role. The situation can be remedied surgically by extirpation of the cataractous lens. The limitations of cataract surgery have stimulated experimental cataract research in laboratory animals and epidemiological studies to determine the incidence, prevalence and risk factors for the development of cataract so as to focus on the preventive aspects of cataract (Gupta *et al.*, 1997a)

Cataract was induced *in vitro* with glucose at a concentration of 55 mM in aqueous humor media and incubated for 72 h at room temperature (Langade 2006). After incubation the lens homogenate was used for the estimation of total protein (TP) content, determination of end products of lipid peroxidation namely malondialdehyde (MDA) and lipid hydroperoxides (LH), enzymatic antioxidants like superoxide dismutase (SOD), catalase (CAT), glutathione reductase (GSSH), peroxidase (Px), glutathione peroxidase (GPx), and non-enzymatic antioxidant reduced glutathione (GSH).

Lipid peroxidation is an autocatalytic process, which is a common cause of cell deth (Bandhyopadhyay et al., 1999). In order to elucidate the protective mechanism of the leaves of Tamarindus indica, glucose-induced goat lens was examined for lipidperoxide levels. Decomposition of lipid peroxides initiate the chain reactions that produce reactive carbonyl compounds. The by-products of lipid peroxidation are the toxic compounds malondialdehyde (MDA) and lipid hydroperoxides (LH) whose involvement in cataractogenesis has been suggested, mainly due to its cross linking ability. Lens MDA may be the result of lipid peroxidation of the lens cell membrane or may represent the consequence of its migration from the readily peroxidizable retina or from the central compartment. In our studies, glucose-induced goat lenses showed an increase in malondialdehyde and lipid hydroperoxide levels in lens. Incubation at different concentrations (100 & 200 of extract of Tamarindus simultaneously with glucose (55 mM) for 72h caused a significant (P<0.01) decrease in the lens malondialdehyde and lipid hydroperoxides and an increase in total protein level. This effect was almost similar to the vitamin E treated group.

ACKNOWLEDGMENT

It is a moment of gratification and pride to look back with a sense of contentment at the long travelled path, to be able to recapture some of the fine moments, to be able to thank infinite number of people, some who were with me from the beginning some who joined me at some stage during the journey, whose kindness, love and blessing has brought this day, I wish to thank each one of them with all my heart.

First and foremost, I bow before the almighty Goddes, Saraswati and thank her for showering her blessings on me and giving me the strength to carry out the present work with utmost dedication and enthusiasm.

Words seem insufficient to express my deep sense of gratitude to beloved and respected guide Dr. Lahu D. Hingane sir, Aditya Pharmacy College, Beed for their excellent guidance, critical supervision, keen interest and continuous encouragement throughout this study. Their active guidance helped me to develop skills and insight in research and scientific presentation. I am

thankful for all the affection, enthusiasm, care and moral support extended to me right from the onset of this uphill task till its completion.

I am heartly thankful to Dr. Lahu D. Hingane Principal of Aditya Pharmacy College, Beed for providing facilities and congenial environment for carrying out my work.

I am obliged and thankful to teaching staff Miss.Tarkase A S, Miss.Salgar P A and non-teaching staff Mr. Narnale S.R., Mr. Pawar V.H. et al. of APC family for their timely help, encouragement, boosting my confidences in the progress of my academics.

It gives me immense pleasure to record my sincere thanks to all my M. Pharm friends for helping me in carrying out this work. It gives me immense pleasure to record my sincere thanks to my senior and juniors.

Finally, the corner of my heart, I express my heartfelt gratitude to my beloved "Parents" and my family members for their initiation, constant source of inspiration, Sacrifice and encouragement to make this work a valuable treasure for me. I am very thankful them for their huge efforts towards my education right from my kinder garden like a strong pillar throughout my life.

Last but not the least, I express my gratitude and apologize to everybody whose contributions, I could not mention in this page.

REFERENCE

- [1] (Bandyopadhyay U, Das D and Banerjee RK (1999). Reactive oxygen species oxidative damage and pathogenesis. Current Science 77: 658-666.
- [2] Bensch KG, Fleming JE and Lohmann W (1985). The role of ascorbic acid senile cataract. Journal of Biochemistry 82: 7193-7196.
- [3] Beydemir S, Kulacoglu DN, Ciftci M and Kufrevioglu OI (2003). The in vitro effect of Dexamethasone on Sheep lens Glucose-6-Phosphate Dehydrogenase. Turkish Journal of Chemistry 27: 601-608.
- [4] Chandorkar AG, Albal MV, Bulakh PM and Muley MP (1981). Lens organ culture. Indian Journal of Ophthalmology 29: 151-154.
- [5] Chylack LT and Kinoshita JH (1969). A biochemical evaluation of a cataract induced in a

- high-glucose medium. Investigative Ophthalmology 8: 401-412.
- [6] Daniyan SY and Muhammed HB (2008). Evaluation of the antimicrobial activities and phytochemical properties of extracts of Tamaridus indica against some diseases causing bacteria. African Journal of Biotechnology 7: 2451-2453.
- [7] Datiles M, Fukui H, Kuwabara T and Kinoshita JH (1982). Galactose cataract prevention with sorbinil, an aldose reductase inhibitor: a light microscopic study. Investigative Ophthalmology and Visual Science 22: 174-179.
- [8] Doughari JH (2006). Antimicrobial activity of Tamarindus indica Linn. Tropical Journal of Pharmaceutical Research 5: 597-603.
- [9] Gupta SK (2004). Drug screening methods, 1st ed. New Delhi: Jaypee brothers. pp. 333-346.
- [10] Gupta SK, Trivedi D, Srivastava S, Joshi S, Halder N and Varma SD (2003). Lycopene attenuates oxidative stress induced experimental cataract development: An in vitro and in vivo study. Nutrition 19: 794-799.
- [11] Gupta SK, Joshi S, Velpandian T, Awor L and Prakash J (1997a). An update on pharmacological
- [12] prospective for prevention and development of cataract. Indian Journal of Pharmacology 29: 3-10
- [13] Gupta SK, Joshi S, Tandon R and Mathur P (1997b). Topical aspirin provides protection against galactosemic cataract. Indian Journal of Ophthalmology 45: 221225.
- [14] Halliwell B and Chirico S (1993). Lipid peroxidation: its mechanism, measurement, and significance. American Journal of Clinical Nutrition 38: 3-20.
- [15] Hejtmancik JH and Kantorow M (2004). Molecular genetics of age-related cataract. Experimental Eye Reserarch 79: 3-9.
- [16] Heyningen RV and Perie A (1967). The metabolism of naphthalene and its toxic effect on the eye. Biochemistry Journal 102: 842-852.
- [17] Kinoshita JH (1965). Pathways of glucose metabolism in the lens. Investigative Ophthalmology 4: 619-628.
- [18] Kovaceva J, Platenik J, Vejrazka M, Stipek S, Arden T, Cejka C, et al., (2007). Differences in activites of antioxidant superoxide dismutase, glutathione peroxidase and prooxidant xanthine

- oxidoreductase/ xanthine oxidase in the normal corneal epithelium of various mammalian. Physiological Research 56: 105-112.
- [19] Krithikar KR and Basu RD (1981). Indian medicinal plants, 2nd ed. Dehradun: International Book Distributors. pp. 887-890.
- [20] Kyselova Z, Garcia SJ, Gajdosikova A, Gajdosik A and Stefek M (2005a). Temporal relationship between lens protein oxidation and cataract development in streptozotocin-induced diabetic rats. Physiological Research 54: 49-56.
- [21] Kyselova Z, Gaidosik A, Gajdosikova A, Ulicna O, Minalova D, Karasu C, et al., (2005b). Effect of pyridoindole antioxidant stobadine on development of experimental diabetic cataract and on lens protein oxidation in rats: comparison with vitamin E and BHT. Molecular Vision 11: 56-65.
- [22] Langade DG, Rao G, Grime RC, Patki PS and Bulakh PM (2006). In vitro prevention by ACE inhibitors of cataract induced by glucose. Indian Journal of Pharmacology 38:107-110.
- [23] Lee AYW and Chung SSM (1998). Involvement of aldose reductase in naphthalene cataract. Investigative Ophthalmology and Visual Science 39: 193-197.
- [24] Lee AYW and Chung SSM (1999). Contributions of polyol pathway in oxidative stress in diabetic cataract. The FASEB Journal 13: 23-30.
- [25] Lowry OH, Rosenbourgh NJ, Farr AL and Randall RJ (1951). Protein measurement with folin phenol reagent. The Journal of Biological Chemistry 193: 265-275.
- [26] Maiti R, Jana D, Das UK and Ghosh D (2004). Antidiabetic effect of aqueous extract of seed of Tamarindus indica in streptozotocin-induced diabetic rats. Journal of Ethnopharmacology 92: 85-91.
- [27] Micelli-Ferrari T, Vendemiale G, Grattagliano I, Boscia F, Arnese L, Altomare E, et al., (1996). Role of lipid peroxidation in the pathogenesis of myopic and senile cataract. British Journal of Ophthalmology 80:840-843.
- [28] Misra HP and Fridovich I (1972). The role of superoxide anion in the autooxidation of epineohrine and simple assay for superoxide dismutase. The Journal of Biological Chemistry 247: 31703175.

- [29] Mizui T, Kinouchi H and Chan PH (1992). Depletion of brain glutathione by buthionine sulfoximine enhances cerebral ischemic injury in rats. American Journal of Physiology 262: 313-317.
- [30] Muller A, Cadenas E, Graf P and Sies H (1984).

 A novel biologically active seleno-organic compound. Glutathion Peroxidase like activity in vitro and antioxidant capacity of PZ51.

 Biochemical Pharmacology 33: 3235-3239.
- [31] Nieshus WG and Samuelsson B (1986). Formation of MDA from phospholipids arachidonate during microsomal lipid peroxidation. European Journal of Biochemistry 6: 126-130.
- [32] Nijveldt RJ, Nood EV, Hoorn ECV, Boelens PG, Norren KV and Leeuwen PAMV (2001). Flavonoids: a review of probable mechanisms of action and potential applications. American Journal of Clinical Nutrition 74: 418-425.
- [33] Nordberg J and Arner ESJ (2001). Reactive oxygen species, antioxidants and the mammalian thioredoxin system. Free Radical Biology and Medicine 31: 1287-1312.
- [34] Nordmann R (1994). Alcohol and antioxidant systems. Alcohol and Alcoholism 29: 513-52
- [35] Olofsson EM, Marklund SL and Behndig A (2007). Glucoseinduced in CuZn-SOD null lenses: An effect of nitric oxide. Free Radical Biology and Medicine 42: 10981105.
- [36] Osakabe N, Yamagishi M, Natsume M, Yasuda A and Osawa T (2004). Ingestion of Proanthocyanidins derived from Cacao inhibits diabetes- induced cataract formation in rats. Experimental Biology and Medicine 229: 33-39.
- [37] Ozmen D, Ozmen B, Erkin E, Guner I, Habil S and Bayindir O (2000). Lens superoxide dismutase and catalase activities in diabetic cataract. Turkish Journal of Endocrinology and Metabolism 1: 1-4.
- [38] Racker E (1955). Glutathione reductase from bakers' yeast and beef liver. The Journal of Biological Chemistry 217: 855-866.
- [39] Raju TN, Kanth VR and Reddy PUM (2007). Influence of kynurenines in pathogenesis of cataract formation in tryptophan –deficient regimen in Wistar rats. Indian Journal of Experimental Biology 45: 543-548.

- [40] Ramos A, Visozo A, Piloto J, Garcia A, Rodjiguez CA and Rivero R (2003). Screening of antimutagenicity via antioxidant activity in cuban medicinal plants. Journal of Ethnopharmacology 87: 241-246.
- [41] Ray G and Husain SA (2002). Oxidants, antioxidants and carcinogenesis. Indian Journal Experimental Biology 40: 1213-1232.
- [42] Robertson JM, Donner AP and Trevithick JR (1991). A possible role for vitamin C and E in cataract prevention. The American Journal of Clinical Nutrition 53: 346S-351S.
- [43] Sato S (1993). Aldose reductase the major protein associated with naphthalene dihydrodiol dehydrogenase activity in rat lens. Investigative Opthamology and Visual Science 34: 3172-3178.
- [44] Schmitt C and Hockwin O (1990). The mechanism of cataract formation. Journal of Inherent Metabolic Disorder. 13: 501-508.